

CENTRO DE INVESTIGACION EN SANIDAD ANIMAL (CISA-INIA)

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SOP/CISA/ASF/PCR/2/

STANDARD OPERATION PROCEDURE FOR THE
DETECTION OF AFRICAN SWINE FEVER VIRUS (ASFV) BY
THE OIE REAL-TIME POLYMERASE CHAIN REACTION (PCR)
1, King *et al.*, 2003.

CONTENTS

1.	PURPOSE
2.	SCOPE
3.	REFERENCES
3.1.	DOCUMENTS USED IN THE PROCEDURE REDACTION
4.	BACKGROUND INFORMATION
4.1.	ABBREVIATION
4.2.	BACKGROUND
5.	PROCEDURE DESCRIPTION
5.1.	EQUIPMENT AND MATERIALS
5.2.	PREPARATION
5.3.	METHODOLOGY
5.4.	ANALYSIS AND INTERPRETATION OF THE RESULTS
5.5.	CRITICAL POINTS
5.6.	SECURITY MEASURES

1. PURPOSE

The purpose of this procedure is to rapidly detect the specific presence of African Swine Fever Virus (ASFV) DNA in porcine clinical material by the real-time polymerase chain reaction (PCR) technique.

Currently this technique is included as OIE - prescribed real time PCR test in the Chapter 3.8.1 of the Manual of Diagnostic Tests and Vaccines for Terrestrial Animals 2019 Edition.

2. SCOPE

This procedure is applied to the ASFV DNA extracted following the procedure described in the **SOP/CISA/ASF/DNA EXTRACTION/1** (“*Standard operating procedure for the extraction of African Swine Fever Virus (ASFV) DNA*”) in any kind of porcine clinical sample such as EDTA-blood, serum and tissue homogenates and in cell culture supernatants. It is particularly useful for identifying ASFV DNA in porcine tissues that are unsuitable for virus isolation or antigen detection, because they have undergone putrefaction, or when there is good reason to believe that virus may have been inactivated before samples are received in the laboratory. PCR technique is highly sensitive, and its detection limit is below one infectious viral particle.

3. REFERENCES

3.1. DOCUMENTS USED IN THE PROCEDURE REDACTION

For the elaboration of this procedure it has been used all the internal procedure included in the INIA-CISA quality system accredited under the UN-EN ISO/IEC17025:2017

In addition as a basic reference for the preparation of this procedure it has been used the following documents:

1. AFRICAN SWINE FEVER. Manual of Diagnostic Tests and Vaccines for Terrestrial Animals (mammals, birds and bees). CHAPTER 3.8.1. OIE, 2019
http://www.oie.int/fileadmin/Home/eng/Health_standards/tahm/3.08.01_ASF.pdf
2. Gallardo C, Nieto R, Soler A, Pelayo V, Fernández-Pinero J, Markowska-Daniel, Pridotkas G, Nurmoja I, Granta R, Simón A, Pérez C, Martín E, Fernández-Pacheco P, Arias M. Assessment of African Swine Fever Diagnostic Techniques as a Response to the Epidemic Outbreaks in Eastern European Union Countries: How To Improve Surveillance and Control Programs. J Clin Microbiol. 2015 Aug;53(8):2555-65.
3. D.P. King, S.M. Reid, G.H. Hutchings, S.S. Grierson, P.J. Wilkinson, L.K. Dixon, A.D.S. Bastos, T.W. Drew. 2003. « Development of a TaqMan PCR assay with internal amplification control for the detection of African Swine Fever Virus”. J. Virol. Methods, 107: 53-61.

ASF REVIEWS:

- Procedure of samples processing for African swine fever (ASF) diagnosis (**SOP/CISA/SAMPLE/1**).
- Procedure for the extraction of African Swine Fever Virus (ASFV) DNA (**SOP/CISA/ASF/DNA EXTRACTION/1**).
- Procedure Quantifast Probe PCR Handbook Qiagen.

4. BACKGROUND INFORMATION

4.1. ABBREVIATION

ASF: African swine fever
ASFV: African swine fever virus
DNA: Deoxyribonucleic acid
E+: ASFV positive extraction control
E-: ASFV negative extraction control
R+: ASFV DNA reaction positive control

R-: ASFV DNA reaction negative control
PCR: polymerase chain reaction

4.2. BACKGROUND

Polymerase chain reaction (PCR) is a molecular genetic technique which allows the specific detection of ASFV DNA by enzyme-based amplification of a short viral genome fragment defined by a specific primer set. Under controlled conditions, multiple copies of DNA are generated by the action of the **DNA polymerase enzyme**, that add complimentary **deoxynucleotides (dNTPs)** to a piece of DNA known as the "template". Real-time PCR is an advanced amplification method, which allows the automated detection of the amplified product, reducing the risk of carry-over contamination with increased specificity and in most cases, even sensitivity.

PCR method requires a first step of viral DNA extraction from the original material to be analysed, which will be the template for the PCR. In real-time PCR, the appearance of amplified product is monitored continuously, in special equipments, with the incorporation in the reaction mix of a fluorescent dye that will give a fluorescence signal in a proportional way to the amplicon accumulation. By determination of fluorescence signal intensity in each amplification cycle, a sigmoid-shaped curve, that represents the amplicon appearance along the PCR, will be obtained.

The described ASFV real-time PCR method uses a primer set and a specific TaqMan probe directed to a highly conserved region of the viral genome, VP72, which ensure the detection of a wide range of ASFV isolates, **belonging to all the 24 known virus genotypes**. The primers amplify a DNA fragment of **250 bp, from nucleotide position 2041 to 2290 of the complete VP72 gene sequence** of the reference strain BA71V (*GenBank accession no. ASU18466*). TaqMan probe employed for amplified product detection is labelled with a reporter at 5' end [6-carboxy-fluorescein (FAM)] and a quencher at 3' end [6-carboxy-tetramethyl-rhodamine (TAMRA)].

PCR is a rapid method, which can be performed in less than four hours, and highly sensitive, allowing the viral detection even before the appearance of clinical symptoms

in the infected animals. PCR is the tool of choice in case of **hyperacute, acute or subacute infection with ASF**.

5. PROCEDURE DESCRIPTION

Method Validation.

Since 2003 the EURL for ASF (INIA-CISA) organize the Annual interlaboratory comparison test (ILCT) to evaluate the ASF diagnostic techniques available at the EU National reference laboratories (NRLs). The ILCT comprises a panel of blind samples to be tested using all the methods routinely employed at the NRLs. This procedure has been fully validated over the last ILCT editions since the technique was developed.

5.1. EQUIPMENT AND MATERIALS

MATERIALS

- Freezer <-10°C.
- Freezer ≤-70°C.
- Fridge 4±3°C.
- Microcentrifuge for eppendorf tubes.
- PC connected to thermocycler.
- Real-time thermocycler [MX3005P, Stratagene or similar characteristics].
- Single channel pipette 1-10µl.
- Single channel pipette 10-100µl.
- Single channel pipette 10-200µl.
- Single channel pipette 200-1000µl
- Tube racks.
- Vortex

DISPOSAL MATERIAL

- Latex or nitrile gloves.
- Microcentrifuge tubes of volumes 0.2 (optical quality), 0.5, 1.5, and 2 ml, sterile.

- Microcentrifuge tubes of volume 0.5 ml, amber colour.
- Sterile filter tips.

⇒ Nuclease-free sterile H₂O, PCR grade.

REAGENTS.

- **Positive and negative controls:** the following controls must be included in each PCR run:
 - ⇒ **E+→ ASFV positive sample target control for the extraction:** ASFV positive sample (serum, EDTA-blood, 1/10 tissue homogenates or culture supernatants) diluted in negative sample. It's highly recommended that the positive control is in the detection limit of the technique to track the yield of the DNA extraction procedure. *Store <-10°C in aliquots. Expiry date: 6 months.*
 - ⇒ **E-→ negative sample control for the extraction:** distilled water which is included during the extraction process to exclude contaminations.
 - ⇒ **R+→ASFV positive DNA target control for the reaction:** ASFV positive DNA. It's highly recommended that the positive control is in the detection limit of the technique to track the yield of the PCR procedure *Store <-10°C in aliquots. Expiry date: 6 months.*
 - ⇒ **R-→ negative DNA target control for the reaction:** distilled water which is included during the PCR process to exclude contaminations.
- **Amplification reagents:**
 - ⇒ **QuantiFast Probe PCR kit, commercially available from Qiagen (Ref. 204352, 100 reactions; ref. 204354, 500 reactions).** *Store at <-10°C.*
 - ⇒ **TaqMan probe** at a concentration of 10 pmol/μl: 5'-FAM-CCACGGGAGGAATACCAACCCAGTG-TAMRA-3'. *Store <-10°C in aliquot, kept always out of light. Expiry date: 1 year*
 - ⇒ **Primers** at a concentration of 20 pmol/μl *Store <-10°C in aliquots. Expiry date: 1 year:*
 - primer **King-s** sequence 5'-CTGCTCATGGTATCAATCTTATCGA-3' (forward primer);
 - primer **King-a** sequence 5'-GATACCACAAGATCRGCCGT-3' (reverse primer).

5.2. PREPARATION

The sample preparation is described in the procedures of samples processing and DNA extraction [(SOP/CISA/SAMPLE/1) and (SOP/CISA/ASF/DNA EXTRACTION/1)].

5.3. METHODOLOGY

5.3.1. DNA AMPLIFICATION.

1. Master mix preparation:

In a sterile 1.5 ml amber Microcentrifuge tube prepare the PCR reaction mixtures described below for the number of samples to be assayed (including R+ and R- controls) allowing for at least two extra samples.

Pipetting step	MASTER MIX REAGENTS	1x VOLUME (reaction 20μl)	FINAL CONCENTRATION
1	H ₂ O	6.7μl	
2	Master mix 2X	10 μl	1X
3	Primer King-s 20 μM	0.4μl	0.4 μM
4	Primer King-a 20 μM	0.4μl	0.4 μM
5	TaqMan probe 10 μM	0.5μl	0.25 μM
Master mix volume		18 μl	

Add 18 μl of the PCR reaction mix to the required number of 0.2 ml optical PCR tubes including the positives controls (+R and +E) and the negative controls (-E and -R), adding at least for one additional sample to minimize pipetting mistakes.

2. Sample addition:

- Add 2μl of DNA template to each PCR tube.
- Include **R+ control** (2 μl of ASFV DNA) and **R- control** (2 μl of distilled water)
- After addition of the template, close the reaction tube and spin down the PCR mix. Place all tubes in an automated real-time thermocycler. Run the incubation program detailed below.

3. PCR CYCLE CONDITIONS.

PCR STEP	Temperature	Time	Nº cycles
Activation of TaqGold DNA pol	95°C	3 min	1x
DNA denaturation	95°C	10 sec	45 x
Primer annealing/elongation	58°C	30 sec	
<i>Program the fluorescence collection in FAM channel at the end of each cycle.</i>			

5.4. ANALYSIS AND INTERPRETATION OF RESULTS

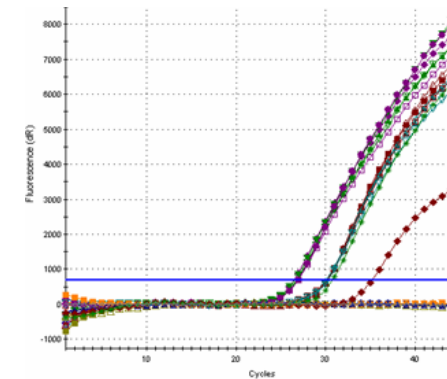
The point where the fluorescence measurement is above the background signal and goes into detectable level is called the cycle threshold (Ct). This will be the starting fluorescence point to consider a sample as positive. The Ct value is inversely proportional to the starting amount of DNA template present in the reaction mixture that means in the analysed sample. The Ct value is automatically determined by the software of the thermocycler

Assay validation:

The assay will be considered as validated when the E+ and R+ have a Ct value within the range 32±4 and the E- and R- Ct ≥40. Otherwise, the analysis should be repeated

Interpretation of the results:

In a **positive sample**, a sigmoid-shaped amplification curve will be obtained, indicating the cycles number versus read fluorescence level, where the Ct value will be under 40. A **negative sample** will maintain the fluorescence profile under background fluorescence level and the equipment will not report any Ct value (see figure). Therefore, a negative sample will show a Ct value ≥40.



5.5. CRITICAL POINTS

Because of PCR is a highly sensitive technique, the most critical point along all the analysis procedure is the considerable risk of carry-over contamination, and the false positive results that could be obtained in this situation. The contamination could be due to the ASFV itself present in the positive analysed samples or in the positive controls included in the DNA extraction procedure; also, it could be due to ASFV DNA obtained after amplification of a previous PCR. **It is mandatory that personnel working on PCR follow and carry out some strict work rules in order to minimize the contamination risk associated to PCR technique:**

- All steps of sample analysis by PCR should be performed in separate locations, using equipment and material specific for each one: sample preparation, DNA extraction, PCR mix preparation, and removal of PCR products.
- Personnel must work always with clean nitrile or latex gloves in the PCR laboratory.
- Whenever personnel goes into a different PCR area, should be remove the gloves and take clean ones.
- The material will be of exclusive use for the PCR procedure step in which is located/labelled.
- Use a new pipette tip each time that a tube containing any sample or DNA is manipulated.

CENTRO DE INVESTIGACIÓN EN SANIDAD ANIMAL (CISA – INIA)	DETECTION OF AFRICAN SWINE FEVER VIRUS (ASFV) BY THE OIE REAL-TIME POLYMERASE CHAIN REACTION (PCR) 1, King <i>et al.</i> , 2003 Rev 6 (2021)	SOP/CISA/ASF/PCR/2
		Page 6 of 7

- Tubes containing amplified product should never be opened and manipulated in other laboratory distinct to that exclusively assigned to their analysis by electrophoresis, where they will be discarded.

5.6. SECURITY MEASURES

- Read and follow carefully the complete procedure.
- Keep reagents to the appropriate temperature before and after use.
- Do not pool reagents or instructions from different kits.
- Avoid any contamination of reagents.
- Do not use any reagent after its expiration date has passed.
- Do not eat, drink or smoke in the laboratory.
- Do not pipette by mouth.
- Wear always protective disposable nitrile or latex gloves.
- Probes used for amplified product detection are highly sensitive to light, so they should be manipulated for the minimum time, and must be kept always out of light (it is recommended to use amber colour tubes to keep probes stock and to prepare reaction mix).

